

Current Biology

The Iceman's Last Meal Consisted of Fat, Wild Meat, and Cereals

Highlights

- The last meal of the Iceman, a European Copper Age mummy, was reconstructed
- Our multipronged approach deciphers the meal composition and food processing
- His high-fat diet was supplemented with wild meat and cereals

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In Brief

Maixner et al. report the dietary reconstruction of the Iceman's last meal using a combined multi-omics approach. The stomach content analysis of the 5,300-year-old glacier mummy shows that the Iceman's diet preceding his death was a mix of carbohydrates, proteins, and lipids, well adjusted to the energetic requirements of his high-altitude trekking.



The Iceman's Last Meal Consisted of Fat, Wild Meat, and Cereals

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SUMMARY

The history of humankind is marked by the constant adoption of new dietary habits affecting human physiology, metabolism, and even the development of nutrition-related disorders. Despite clear archaeological evidence for the shift from hunter-gatherer lifestyle to agriculture in Neolithic Europe [1], very little information exists on the daily dietary habits of our ancestors. By undertaking a complementary -omics approach combined with microscopy, we analyzed the stomach content of the Iceman, a 5,300-year-old European glacier mummy [2, 3]. He seems to have had a remarkably high proportion of fat in his diet, supplemented with fresh or dried wild meat,

cereals, and traces of toxic bracken. Our multi-pronged approach provides unprecedented analytical depth, deciphering the nutritional habit, meal composition, and food-processing methods of this Copper Age individual.

RESULTS AND DISCUSSION

Human evolution is closely linked to dietary changes and the development of new food-processing techniques. This is clearly observed in the transition from hunter-gatherer lifestyle to agriculture, which gave rise to cultivation of crops, animal husbandry, and permanent settlements [1]. The more stable availability of food boosted the growth of Neolithic European populations [4, 5]. However, changes in diet had drawbacks



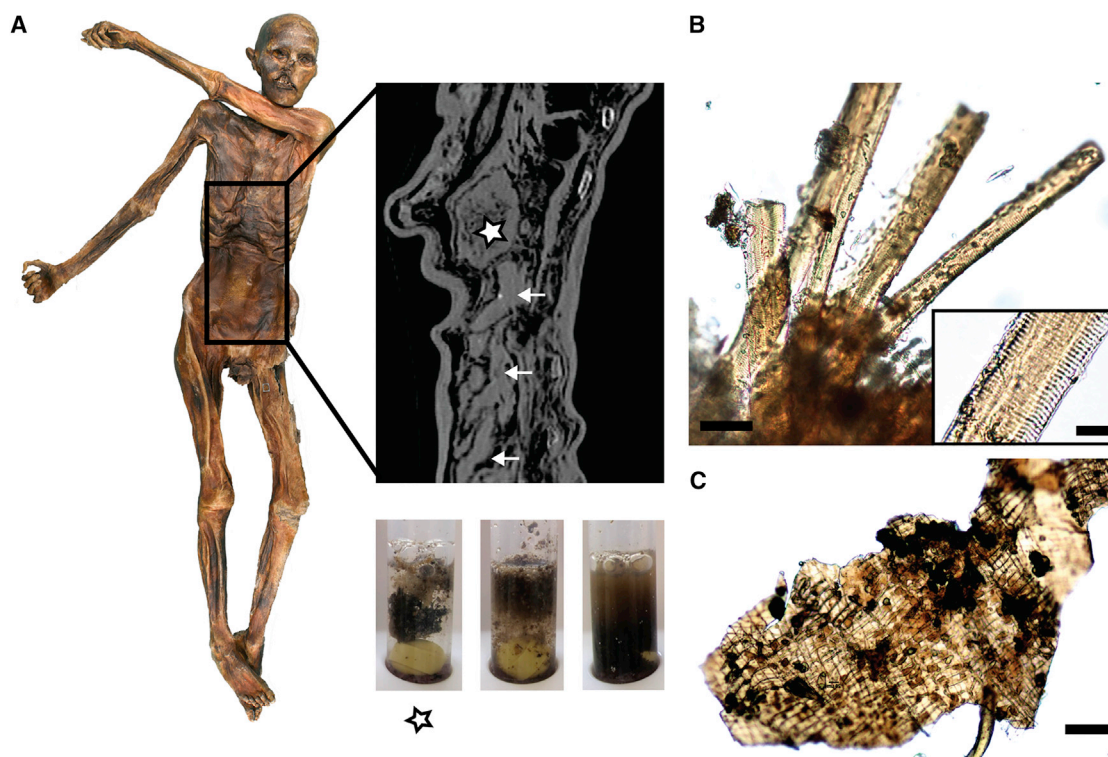


Figure 1. The Iceman's Gastrointestinal Tract

(A) Gastrointestinal (GI) tract preservation and content texture. The radiographic image shows the completely filled stomach (asterisk) and the intestinal loops of the lower GI tract (arrows). Content samples of the stomach (left, asterisk) and of two different sites in the lower GI tract (middle, right) that were re-hydrated in phosphate buffer are shown below the radiographic image.

(B) Animal muscle fibers detected in the stomach content using light microscopy. Scale bar, 50 μm. The black box contains a zoomed-in view of one muscle fiber (scale bar, 20 μm).

(C) Plant tissue detected in the stomach content using light microscopy. Scale bar, 50 μm. See also [Figure S1](#) for additional plant and animal remains detected in the Iceman intestinal contents.

for health, such as increased rates of caries [6]. Added to this, permanent large settlements combined with the adoption of agriculture promoted the spread of density-dependent infectious diseases [7]. Considering the impact of dietary habits on human evolution and health, the reconstruction of past human subsistence appears to be vital to our understanding of past societies. Stable carbon ($^{13}\text{C}/^{12}\text{C}$) and nitrogen ($^{15}\text{N}/^{14}\text{N}$) isotope analyses of bone collagen have been widely used to study the dietary adaptations of Neolithic populations [8], lipid analysis on pottery residues dated the onset of milk use in Neolithic Europe [9], and numerous archaeological studies describe the occurrences of wild and domesticated animal and plant remains as indirect measures of past diets [10]. All these studies provide insights into major dietary changes during the Neolithic period. However, specific details on past diets such as the exact meal composition or food processing are missing. Knowledge of daily dietary habits of our ancestors is pertinent as food type, intake quantity, and food-processing techniques influence human metabolism and health.

A recent radiological re-examination of the Iceman, a 5,300-year-old European natural ice mummy, identified his completely filled stomach (Figure 1A) [2]. The well-preserved stomach content still contains ancient endogenous biomolecules as

demonstrated from the complete reconstruction of the Iceman's *Helicobacter pylori* genome [11]. Previous isotopic analysis ($^{15}\text{N}/^{14}\text{N}$) of the Iceman's hair suggested first a vegetarian lifestyle [12, 13] which was later, after careful re-examination of the data, changed to an omnivorous diet [14]. Further analyses of lower intestinal tract samples of the Iceman confirmed that he was omnivorous, with a diet consisting of both wild animal and plant material. Among the plant remains, there were cereals, pollen grains of hop-hornbeam, and fragments of bracken and mosses [14–20]. The detection of the Iceman's stomach content with its pristine yet undigested food mix, provides the unique opportunity to fully reconstruct a Copper Age meal.

The study material included 11 gastrointestinal (GI) content biopsies of the stomach and small and large intestines (Figure 1A; [Data S1](#)). Using microscopy and a combined multi-omics approach targeting different biomolecules (ancient DNA, proteins, metabolites, and lipids) (see [STAR Methods](#)), we determined the exact composition of the Iceman's diet preceding his death and present evidence how people during the Copper Age processed food.

Initial macro- and microscopic analyses showed that, compared to the small and large intestine contents, samples of the stomach content still contain compact pieces of food that

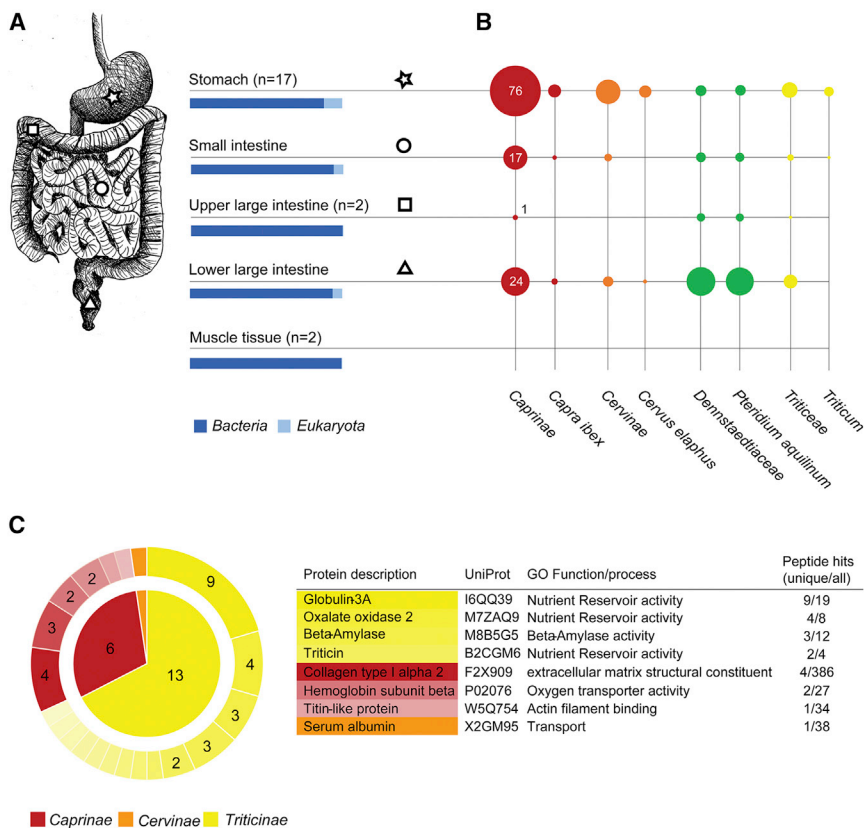


Figure 2. Metagenomic and Proteomic Analysis

(A) GI tract samples included in the metagenomic approach. The blue-bar diagrams below the sample description display the read distribution between the two kingdoms *Bacteria* and *Eukaryota* in selected shotgun datasets (for details, please refer to [STAR Methods](#) and [Data S1](#)). The number of shotgun datasets included in all further metagenomic analyses is provided in brackets.

(B) Most abundant taxa detected in the intestinal content shotgun datasets. The circle size corresponds to the number of unambiguously assigned mitochondrial and chloroplast reads per million metagenomic reads.

(C) Plant and animal proteins detected in the stomach content. The inner circle displays the number of identified proteins for the taxa *Caprinae*, *Cervinae*, and *Triticinae*. The outer circle highlights the number of unique peptides present for protein identification. The table summarizes details about the proteins detected with the highest unique peptide hits. See also [Figures S2, S3, and S4](#) for a taxonomic overview of the plastid reads, the phylogenetic assignments, damage pattern, and DNA-barcode analysis. [Data S1](#) contains details to the datasets and the plastid mapping statistics. [Data S2](#) provides additional information to the identified peptide hits.

display a hydrophobic “fatty-like” character ([Figure 1A](#)). The most prevalent tissue types in the stomach content samples are animal muscle fibers arranged mostly in bundles ([Figures 1B and S1](#)) and plant fragments identified as bran and glumes belonging to the *Triticum/Secale* type ([Figures 1C and S1](#)). The striated muscle fiber structures are characteristic for cardiac and skeletal muscle tissue ([Figure S1B](#)) [21]. Further microscopic analysis of the plant micro-remains identified cereal pollen from the *Triticum*-type as well as airborne arboreal pollen and an abundance of spores and sporangia from the bracken fern *Pteridium aquilinum* ([Figure S1J](#)).

Metabolite, glycan, stable isotope, and elemental analyses confirm the omnivorous character of the Iceman’s last meal ([Data S2](#)). Metabolic compounds indicate the presence of ruminant fat or dairy products (phytanic acid) and support the existence of whole-grain cereals (azealic acid) in the Iceman’s diet. In addition, gamma-terpinene that occurs in coriander oil, lemon oil, and other essential oils suggest the usage of herbs. The most abundant elements found in the stomach content were the nutritional minerals iron, calcium, zinc, magnesium, and sodium, consistent with the consumption of red meat or dairy products. Minor concentrations of trace nutrients of chromium, copper, manganese, selenium, molybdenum, and cobalt were also detected. These data suggest that the Iceman’s last meal was well balanced in terms of essential minerals required for good health with no evidence of toxic heavy metals such as lead, cadmium, or arsenic.

To further identify the possible food sources, we analyzed the DNA and protein traces still present in the material. Metage-

nom analysis of the Iceman’s biopsy samples revealed a high number of bacterial reads in all GI tract contents ([Figure 2A](#)). At most, 39% of reads in the Iceman samples were of eukaryotic origin, the majority of which (between 94% and 42%) were identified as fungal. The *Metazoa* and *Viridiplantae* reads, important for the dietary reconstruction, comprised 0.2%–0.7% of all reads ([Figures S2A–S2C](#)). Comparative analysis of all GI shotgun datasets ($n = 21$) against currently available full mitochondrial and chloroplast genomes unambiguously assigned the majority of animal reads to ibex (*Capra ibex*) and red deer (*Cervus elaphus*), and the major proportion of plant reads belong to the species *Pteridium aquilinum* subsp. *aquilinum* and to members of the genus *Triticum* ([Figures 2B and S2D](#)). DNA traces of these four major taxa were detected in all GI content samples. A control dataset from the Iceman’s muscle tissue was negative ([Figure S2D](#)). The unambiguously assigned reads were aligned to modern reference plastid genomes, and all animal- and plant-derived reads show damage patterns indicative of ancient DNA ([Figure S3](#)) [22]. Subsequent phylogenetic assignment of the reconstructed plastid genomes confirmed and further extended the metagenomic result ([Figure S3](#)). The *Triticum* reads were more specifically assigned to chloroplast genomes of *Triticum monococcum* (einkorn) and *Triticum urartu* ([Figure S3; Data S1](#)).

Importantly, the presence of ibex and red deer in the stomach contents was independently confirmed in the Molecular Population Genetics Laboratory, Trinity College, Dublin, by sequencing of a mammalian mitochondrial capture ([Data S1](#)). Moreover, PCR-based analysis targeting animal- and plant-DNA barcodes and further assignment of the shotgun data against the DNA

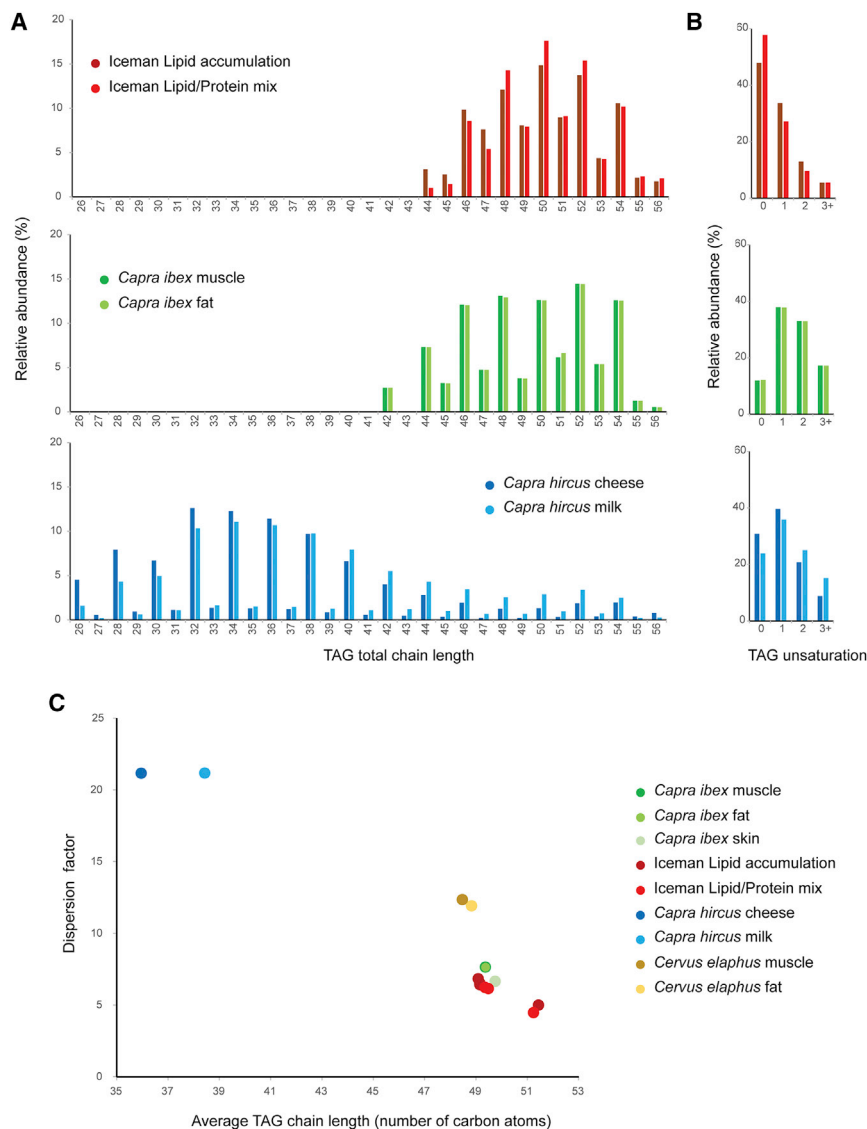


Figure 3. Lipid Analysis of the Iceman's Stomach Content

(A) Triacylglycerol (TAG) distribution in the Iceman's stomach's content in comparison to the TAG distribution in ibex muscle and fat tissue and goat dairy products. Displayed is the relative abundance of TAGs based on their chain length. (B) Saturation level of the TAGs in the stomach content and the fresh tissue samples. The Iceman's stomach TAGs contain a much higher saturated bond content than fresh tissue samples. (C) The dispersion factor (DF) plotted against the average carbon number (M) of each TAG detected in the ancient and modern samples. The DF measures the spread of the TAG distribution. Together with M, DF can be used for discrimination between archaeological artifacts [26]. See also [Data S2](#) for further details to the relative abundance of TAG species and to the mass spectrometry results.

last meal contained whole seeds of the *T. monococcum*/*T. urartu*-like wheat and muscle fibers of ibex. However, due to the lack of diagnostic proteins, we cannot determine which tissues of the red deer were ingested by the Iceman.

Approximately $46\% \pm 19\%$ (w/w) of the stomach content is fat residues. Comparative lipid analysis of the Iceman's stomach content to contemporary wild ruminant muscle and adipose tissue, and to goat dairy products (milk and cheese), using reverse-phase liquid chromatography separation and quadrupole time-of-flight (QToF) tandem mass spectrometry analysis (RP-LC/MSMS) identified triacylglycerols (TAGs) as a major component of the stomach content, with molecular compositions consistent with ruminant muscle/adipose

fat (Figure 3A). The TAGs identified in the Iceman's stomach contain between 44 and 56 total carbon atoms in the fatty acyl chains and resemble the TAG profile of contemporary muscle and adipose tissue. The high level (up to 20%) of odd numbered fatty acyl chains is consistent with TAG profiles from ruminant animals (Data S2) [27]. Short-chain TAG species characteristic of dairy products are absent in the Iceman TAG profile. However, the high level of saturated bonds in the TAGs of the Iceman's stomach content compared to the fresh tissues indicates an already ongoing preferential degradation of unsaturated fatty acids in the ancient specimen (Figure 3B) [28]. Therefore, we cannot fully exclude that short-chain dairy-specific lipid species were degraded, although the identification of di- and poly-unsaturated TAG indicates a still high level of preservation. Furthermore, we have histological evidence for the presence of adipose tissue in the Iceman's stomach content (Figure S1C). Further comparison of the dispersion factor with the average carbon number (M) of each detected TAG in the analyzed samples points to the ibex as potential source of the

Barcode of Life Data dataset (www.boldsystems.org) support the first metagenomic results on a phylogenetic higher hierarchical level (genus, family) (Figure S4). Our analysis of the ancient DNA identified the main components in the Iceman's meal. However, the highly degraded state of this biomolecule excluded any further quantification of certain foods.

Using multistep solubilization and fractionation proteomics, we identified 167 animal and plant proteins in the stomach metaproteome, of which 13 were taxonomically unambiguously assigned to the *Triticinae*, 6 to the *Caprinae*, and 1 protein to the *Cervinae* subfamily (Figure 2C; Data S2). The proteomic hits not only confirm the previous metagenomic results but also provide important information on the food source. The detected *Triticinae* plant proteins are primarily expressed in the endosperm (Globulin 3A, Triticin) [23] and pericarp (Oxalate oxidase 2) [24] of wheat seeds. The *Caprinae* Titin-like protein is part of the sarcomere apparatus in muscle tissue [25]. Thus, we have combined genomic and proteomic evidence that the Iceman's

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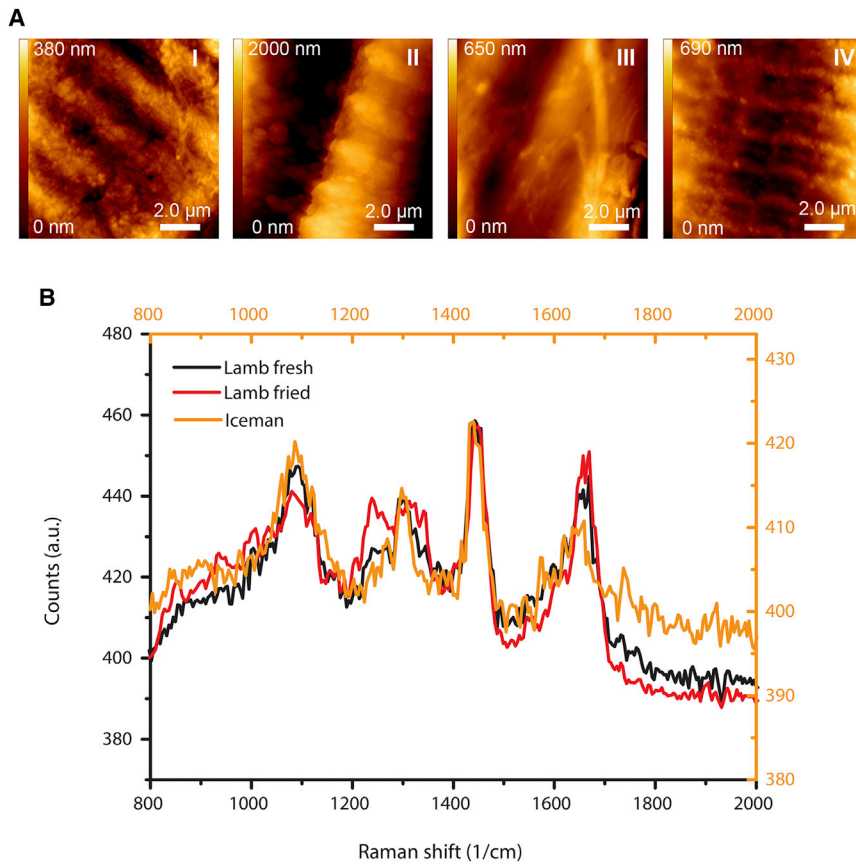


Figure 4. Atomic Force Microscope Images and Raman Spectra of Muscle Fibers from the Iceman's Stomach Content

(A) Fibers obtained from the content of the Iceman's stomach (I) show structures similar to those of recent fibers (II and VI). The typical Z-line is observed in raw (II) and air-dried lamb muscle fibers (IV). After thermal treatment such as frying or cooking of the meat, the sarcomeres disappear (III). The surface of the fibers becomes blurred, and only faint fibrillary structures can be found.

(B) Raman spectra of samples extracted from the Iceman's stomach content show similarities to untreated (fresh) lamb meat spectra. See [Data S2](#) for details to the modern comparative animal samples.

adipose tissue (Figure 3C). Thus, the ibex served the Iceman as food source for both meat and fat.

Charcoal particles detected in a previous study in the Iceman's lower intestine content suggested the involvement of fire in the food processing [16]. We experimentally tested the effect of heat and drying on contemporary animal muscle tissue and compared their structural and chemical composition with the muscle fibers detected in the Iceman's stomach by using atomic force microscopy (AFM) and Raman spectroscopy. High magnification AFM images revealed striated fiber structures in both ancient and fresh or dried contemporary muscle fibers while structural changes of the fibers were observed when the samples were thermally treated (Figure 4A). After frying or cooking, the regular patterns in the meat fibers disappeared, indicating the denaturation of their protein compounds. Complementary confocal Raman spectroscopy measurements support the AFM results and indicate heat-related conformational changes of the molecular composition of the meat fibers (Figure 4B). The Raman spectra of the ancient muscle fibers largely matched the spectra taken of fresh and dried contemporary meat fibers. After heat treatment, the Raman band at about 1,093 cm^{-1} becomes weaker, and bands at around 1,240 cm^{-1} appear that are indicative of heat-induced changes in the secondary structure of the myofibrils [29]. Thus, both the AFM and Raman spectroscopy results suggest that the Iceman consumed either fresh or dried wild meat. Besides, a slow drying or smoking of the meat over the fire would explain the charcoal particles detected previously in the lower intestine content.

Our forensic multi-omics study provides important insights into the general life and nutritional habits of a Copper Age individual in the Alpine area. The microscopic and molecular data presented support the presence of three major components in the Iceman's last meal: fat and game meat from ibex and red deer supplemented with cereals from einkorn. Intriguing is the pervasive presence of bracken particles. Despite of its toxicity, bracken is still consumed today by different indigenous people [30]. However, it is also possible that fern leaves, as it was previously suggested for the moss *Neckera complanata* [20], were used to wrap the food and the spores and sporangia entered the ingesta unintentionally. Unexpectedly, there is a high proportion of fat in the diet. The distribution of triglycerides and their constituting fatty acids is consistent with the consumption of ibex muscle and adipose tissues. The extreme alpine environment in which the Iceman lived and where he has been found (3,210 m above sea level) is particularly challenging for the human physiology and requires optimal nutrient supply to avoid rapid starvation and energy loss [31]. Therefore, the Iceman seemed to have been fully aware that fat displays an excellent energy source. On the other hand, the intake of animal adipose tissue fat has a strong correlation with increased risk of coronary artery disease [32]. A high saturated fats diet raises cholesterol levels in the blood, which in turn can lead to atherosclerosis. Importantly, computed tomography scans of the Iceman showed major calcifications in arteries and the aorta indicating an already advanced atherosclerotic disease state [33]. Both his high-fat diet and his genetic predisposition for cardiovascular disease [34] could have significantly contributed to the development of the arterial calcifications. Finally, we could show that the Iceman either consumed fresh or dried meat. Drying meat by smoking or in the open air are simple but highly effective methods for meat preservation that would have allowed the Iceman to store meat long term on journeys or in periods of food scarcity. In summary, the Iceman's last meal was a well-balanced mix of carbohydrates, proteins, and lipids, perfectly adjusted to the energetic requirements of his high-altitude trekking.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

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SUPPLEMENTAL INFORMATION

Supplemental Information includes four figures and two data files and can be found with this article online at <https://doi.org/10.1016/j.cub.2018.05.067>.

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AUTHOR CONTRIBUTIONS

F.M. and A.Z. conceived the investigation. F.M., D.T., A.C.-G., M.J., M.R.H., R.L.M., A.F., A.K., M.W., R.W.S., O.F., P.D., T.R., R.G., and A.Z. designed the experiments. P.M., P.G., L.E., E.E.-V., M. Samadelli, F.M., and A.Z. were involved in the sampling campaign. F.M., A.C.-G., M.J., B.K.-K., M.R.H., U.K., M. Sartain, N.O., M.T., G.C., V.M., M.P., J.M., S.L., P.R., B.J.K., H.-J.A., and D.B. conducted the experiments. F.M., D.T., A.C.-G., M.J., M.R.H., N.O., M.T., S.L., J.F.B., B.J.K., H.-J.A., and K.O. performed analyses. F.M. wrote the manuscript with contributions from D.T., A.C.-G., M.J.,

M.R.H., A. Paladin, A. Putzer, G.G., U.T., A.N., D.G.B., K.O., R.G., T.R., and A.Z.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Biological Samples		
Iceman gastrointestinal tract content samples and tissues biopsies	[11]	N/A
Chemicals, Peptides, and Recombinant Proteins		
EDTA disodium salt dihydrate	Carl Roth	Cat#8043.2
Chloroform	SIGMA-ALDRICH	Cat#C2432
Chloroform	Fisher Chemical	Cat#C297-4
Chloroform -Isoamylalcohol (24:1, v/v)	SIGMA-ALDRICH	Cat#C0549
Methanol	Carl Roth	Cat#4627.1
Methanol	Fisher Chemical	Cat#A456-4
Acetonitrile	Fisher Chemical	Cat#A955-4
Isopropanol	Fisher Chemical	Cat#A461-4
Cetyltrimethylammonium bromide CTAB	Carl Roth	Cat#9161.3
Polyvinylpyrrolidone-K10	SIGMA-ALDRICH	Cat#PVP10-100G
Tris-HCl	Carl Roth	Cat#9090.2
Acrylamide	Carl Roth	Cat#0189.2
Ammonium peroxydisulphate APS	Carl Roth	Cat#9592.2
TEMED	Carl Roth	Cat#2367.3
Sodium acetate	Carl Roth	Cat#6773.1
Guanidine hydrochloride	Carl Roth	Cat#6069.2
Beta-Mercaptoethanol	Carl Roth	Cat#4227.3
BSA (20 mg/ml)	New England Biolabs	Cat#B9000S
Bst polymerase, large fragment (8 U/μl)	New England Biolabs	Cat#M0275S
dNTPs (2.5mM each)	Thermo Fisher Scientific	Cat#R72501
T4 DNA ligase (5 U/μl)	Thermo Fisher Scientific	Cat#EL0014
Critical Commercial Assays		
Min Elute PCR Purification Kit	QUIAGEN	Cat#28006
Amplitaq Gold mastermix 360	Thermo Fisher Scientific	Cat#4398881
NEBNext End Repair Module	New England Biolabs	Cat#E6050L
Accuprime Pfx Supermix	Thermo Fisher Scientific	Cat#12344040
Deposited Data		
Iceman gastrointestinal tract content shotgun datasets	[11]	ENA: ERP012908
Iceman stomach content enriched dataset	This Study	ENA: PRJEB26465
Iceman stomach content amplicon datasets	This Study	ENA: PRJEB26365
Iceman stomach content proteomics data	This Study	PRIDE: PXD009565
Oligonucleotides		
12419F CAAACTGGGATTAGATACCC	This Study	Thermo Fisher Scientific
12564R YRGAACAGGCTCCTCTAG	This Study	Thermo Fisher Scientific
CBF GCGTACGCAATCTTACGATCAA	[35]	Thermo Fisher Scientific
CBR CTGGCCTCCAATTCATGTGAG	[35]	Thermo Fisher Scientific
trnL-g GGGCAATCCTGAGCCAA	[36]	Thermo Fisher Scientific
trnL-h CCATTGAGTCTCTGCACCTATC	[36]	Thermo Fisher Scientific
rbcl19 AGATTCCGCAGCCACTGCAGCCCCTGCTTC	[37]	Thermo Fisher Scientific
rbclZ1 ATGTCACCACAAACAGAGACTAAAGCAAGT	[37]	Thermo Fisher Scientific

(Continued on next page)

Continued		
REAGENT or RESOURCE	SOURCE	IDENTIFIER
Software and Algorithms		
Imaging Software NIS elements F 3.00	Nikon	N/A
ChromaTOF software version 4.2	LECO	N/A
FastQC	Babraham Bioinformatics	http://www.bioinformatics.babraham.ac.uk/projects/fastqc/
SeqPrep	N/A	https://github.com/jstjohn/SeqPrep
RAPSearch2	[38]	http://rapsearch2.sourceforge.net/
Krona	[39]	https://github.com/marbl/Krona
MEGAN6	[40]	https://ab.inf.uni-tuebingen.de/software/megan6
Burrows-Wheeler Aligner (BWA)	[41]	http://bio-bwa.sourceforge.net/
blastn	[42]	N/A
FastQ Screen	Babraham Bioinformatics	http://www.bioinformatics.babraham.ac.uk/projects/fastq_screen/
SAMtools	[43]	http://samtools.sourceforge.net/
Picard tools	N/A	http://broadinstitute.github.io/picard/
ANGSD	[44]	http://www.popgen.dk/angsd/index.php/ANGSD
MAFFT	[45]	https://mafft.cbrc.jp/alignment/server/
ARB software package	[46]	http://www.arb-home.de/
AdapterRemoval	[47]	https://github.com/MikkelSchubert/adapterremoval
DeDup	N/A	https://github.com/apeltzer/DeDup
mapDamage	[21]	https://ginolhac.github.io/mapDamage/
GATK	[48]	https://software.broadinstitute.org/gatk/
mothur	[49]	https://www.mothur.org/
Trans-Proteomic Pipeline (TPP) software tool suite	[50]	http://tools.proteomecenter.org/wiki/index.php?title=Software:TPP
MassHunter Qualitative Analysis Software Version B.05.01	Agilent	https://www.agilent.com/en/products/software-informatics/masshunter-suite/masshunter/masshunter-software
Other		
BOLD system database	[51]	http://www.boldsystems.org/
Additional metadata for the Illumina shotgun datasets	[11]	http://cube.univie.ac.at/supplements/iceman

CONTACT FOR REAGENT AND RESOURCE SHARING

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Frank Maixner (frank.maixner@eurac.edu).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

The Iceman

The Iceman is one of the oldest human mummies ever found in human history [52]. With his body perfectly preserved throughout millennia in an Alpine glacier (3,210 m above sea level) and his secrets unlocked and shown for the first time to humanity by the melting of ice in 1991, it has been and still is a subject of great interest, fascination and speculations, since the veil of mystery enveloping the circumstances of his death has not yet fallen [3]. The Iceman was murdered when he was 40-50 years old by an arrow that lacerated the left subclavian artery, likely leading to a rapid, deadly hemorrhagic shock [53, 54]. It is not only his historical age and the bounty of prehistoric belongings that make him extremely valuable for scientists, but also how he was preserved for about 53 centuries. The Iceman is indeed an “ice mummy,” so-called for the retention of humidity in his cells [55]. The body tissues contain well preserved biomolecules such as lipids, proteins, and nucleic acids. This feature makes him extremely valuable for modern scientific investigations [11, 34, 56–61]. Initially, the Iceman’s stomach was not found radiologically and was generally assumed

to be empty. During a recent re-examination however, the Iceman's stomach was not only identified radiologically, but also shown to be completely filled [2].

METHOD DETAILS

Sampling

The sampling of the Iceman stomach content took place under sterile conditions at an ambient temperature of 4°C in the Iceman's conservation chamber at the Archaeological Museum of Bolzano, Italy. The samples were immediately stored at –20°C in the ancient DNA laboratory of the Eurac Research - Institute for Mummy Studies. Furthermore, we had access to previously sampled gastrointestinal tract contents and to an Iceman muscle and lung tissue sample, which we used in addition to the extraction blanks as a negative control for our molecular analyses. See [Data S1](#) for additional details to the samples used in this study.

Microscopic analysis

Small pieces of the stomach content (0.5cmx0.5cm) were processed for microscopy in the aDNA laboratory of the Eurac Research - Institute for Mummy Studies, Bolzano. After rehydration in 15mL solution consisting of 5 parts glycerol and 5 parts 4% formaldehyde for 48 h, the samples were fixed for 24 h in 4% formaldehyde and finally stored in 1x phosphate buffered saline solution at 4°C. For the microscopic analysis, small tissue pieces have been embedded with an aqueous mounting medium on a glass slide. In parallel, some fixed content samples were embedded in OCT cryostat embedding medium (Tissue Tek®) and further processed for histology. The embedded specimens were cut on a cryostat in 10 µm thick sections (Leica, CM1950). The cryosections were histochemically counterstained with the lipid stain Sudan III [62]. The images were recorded with a CCD camera (Nikon DS-Fi1) mounted on a light microscope (Nikon Eclipse E600), by using the imaging software NIS elements F 3.00. Additionally, ancient animal meat fibers detected in the stomach content were sent to the Center of Smart Interfaces at the TU Darmstadt, Germany, where the coarse topography of meat fibers was analyzed by laser scanning microscope (LSM) images (VK 8710, Keyence, Osaka, Japan).

Metabolite analysis

Primary metabolism was assessed using GC-TOF MS. 2 mg fresh weight of stomach contents were extracted using 1 mL of degassed and pre-cooled (–20°C) acetonitrile/isopropanol/water solvent at the volume ratio 3:3:2. After centrifugation, the supernatant was removed and evaporated to complete dryness. A clean-up step with 500 µL acetonitrile/water (1:1) removed membrane lipids and triglycerides and after centrifugation. To the dried extracts, C8-C30 fatty acid methyl esters were added as internal standards. Samples were derivatized by 10 µL methoxyamine hydrochloride in pyridine (20 mg/mL) at 30°C for 90 min, followed by the addition of 90 µL *N*-methyl-*N*-trimethylsilyltrifluoroacetamide at 37°C for 30 min. 0.5 µL of sample was injected at 50°C (ramped to 250°C by 12°C/s and held for 3 min) in splitless mode in a Gerstel cold injection system with multi-baffled glass liners. Metabolites were separated using an Agilent 6890 GC system (Santa Clara, CA, USA) equipped with a Rtx5Sil-MS column (30 m length x 0.25 mm i.d.) with a 0.25 µm 95% dimethyl/ 5% diphenyl polysiloxane film and additional 10 m integrated guard column (Restek, Bellefonte, PA, USA). Chromatography was performed at 1 mL/min helium from 50°C to 330°C with 20°C/min. Mass spectra were acquired on a Leco Pegasus IV time of flight mass spectrometer (St. Joseph, MI, USA) with electron ionization at 70 eV at 250°C. Masses were detected between *m/z* 85-500 at 17 spectra/s and 1800 V detector voltage. ChromaTOF software version 4.2 was used for data pre-processing including baseline subtraction and automatic mass spectral deconvolution and peak detection. The resulting chromatograms were further processed using the metabolomics BinBase database and spectra were further manually investigated by the Fiehnlib libraries and the NIST12 mass spectral library. Please refer for the list of metabolites identified in the Iceman stomach content to [Data S2_Metabolomics](#).

Glycan analysis

Glycan analysis for the Iceman stomach content was performed at Asia-Pacific Glycomics Reference Site (AGRS) in Daejeon, Republic of Korea. Two stomach content samples of 0.021 g and 0.023 g, respectively were ground with 300 µL 70% EtOH into tissue grinder. High molecular weight materials including glycoproteins were enriched from ground samples by mini dialysis unit (10,000 MWCO). Supernatant was transferred to micro-centrifuge tube and denatured by mixture of NH₄HCO₃ and DTT (95°C, 2min). *N*-glycans were enzymatically released from denatured glycoproteins by PNGase F (37°C, overnight) and these were further purified and enriched using solid phase extraction with a porous carbon cartridge. *N*-glycans released from the Iceman stomach content were comprehensively characterized by nano LC PGC-chip/Q-TOF MS and tandem MS (Agilent 6530). Glycan candidates were selected from the mass-to-charge values extracted in raw LC/MS data based on monosaccharide combinations using hexose, *N*-acetyl hexoamine, fucose, *N*-acetyl neuraminic acid, and xylose. *N*-glycan compositions were further confirmed and determined by compositional analysis using tandem MS. In order to determine relative amounts of glycans, individual glycan compound was integrated using the ion count corresponding to individual peak and then normalized by total ion counts to obtain the relative abundance of each glycan. Additionally, isomers derived from PGC-LC analysis were combined in a single composition prior to quantitative analysis. [Data S2_Glycomics](#) lists all identified glycan structures.

Stable isotope analysis

The preparation of the stomach content (EURAC ID 1054) for the stable isotope analysis was conducted in the laboratory at the Department of Physical Anthropology, Institute of Forensic Medicine at University of Bern. Therefore, a sample of 1g was dissolved in ddH₂O and freeze-dried for 47h. After homogenization by a ball triturator (Retsch), approx. 5.5mg of the lyophilized stomach content was weighed into tin capsules and shipped to IRMS at Isolab GmbH, Schweitenkirchen, Germany for measurement. Stable isotope ratios of carbon (¹³C/¹²C), nitrogen (15N/14N) and sulfur (34S/32S) were analyzed by isotope ratio mass spectrometry and the mean of six measurements was calculated to avoid measurement errors due to potential inhomogeneous material. All data are presented in δ-notation in per mil (‰) according to international standards for carbon (Vienna Pee Dee Belemnite, VPDB), nitrogen (Ambient Inhalable Reservoir, AIR) and sulfur (Canyon Diablo Troilite, CDT) [63–65]. An internal standard was used for determination of the analytical error. It amounted to ± 0.1‰ for δ13C, ± 0.2‰ for δ15N and ± 0.3‰ for δ34S. For details for the stable isotope values of the Iceman stomach content please refer to the [Data S2_Stable isotope analysis](#).

Elemental analysis of the Iceman stomach content

An Agilent Technologies 7500cx inductively coupled plasma-mass spectrometer (ICP-MS) (Agilent Technologies, Mulgrave, Australia) was used with sample introduction via a micromist concentric nebuliser (Glass Expansion, West Melbourne, Australia) and a Scott type double pass spray chamber cooled to 2°C. The sample solution and the spray chamber waste were carried with the aid of a peristaltic pump. ICP-MS extraction lens conditions were selected to maximize the sensitivity of a 1% HNO₃:HCl solution containing 1ng ml⁻¹ of Li, Co, Y, Ce and Tl. Helium was added into the octopole reaction cell to reduce interferences. Calibration curves were constructed and the results analyzed using Agilent Technologies MassHunter software. A certified calibration standard containing Li, Be, B, Na, Mg, Al, Ca, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Sr, Mo, Ag, Cd, Sb, Ba, La, Eu, Ho, Yb, Tl, Pb, Bi, Th, U and P as well as a Hg standard was obtained from Choice Analytical, Thornleigh, Australia. Calibration curves were constructed from seven standards at 0, 1, 10, 50, 100, 500, and 1000 ng/mL. Baseline nitric acid (HNO₃) hydrochloric acid (HCl) and hydrogen peroxide (H₂O₂) were purchased from Choice Analytical, Thornleigh, Australia. Two stomach content samples of 0.05915 g and 0.0279 g were each digested in 2 mL HNO₃, 1 mL HCl and 1 mL H₂O₂ in a Milestone MLS 1200 microwave digester (Milestone, Sorisole, Italy), and diluted to approximately 50 g (accurately measured on an analytical balance) before analysis. A digestion blank was subtracted from each sample. The average concentrations of the elements detected in the Iceman's stomach is listed in the [Data S2_Elemental analysis](#).

DNA extraction, library preparation and Illumina sequencing

The datasets used for the metagenomic dietary analysis have been published in our previous study on the Iceman's *Helicobacter pylori* genome. In the following sections, all major molecular steps will be briefly outlined once more. For methodological details, however, please refer to the publication of Maixner et al. [11]. The molecular analyses were conducted at the ancient DNA Laboratory of the Eurac Research - Institute for Mummy Studies, Bolzano, Italy, and in the ancient DNA facility of the Institute of Clinical Molecular Biology, Kiel University, Germany. Sample preparation and DNA extraction was performed in pre-PCR areas dedicated to ancient DNA procedures. DNA was extracted from the gastrointestinal tract samples (250 mg) and from lung tissue (50 mg) using a chloroform-based DNA extraction method according to the protocol of Tang et al. [66]. DNA extraction from the muscle tissue (100 mg) was performed with a magnetic-bead based technology using the Biorobot®-EZ1 (QIAGEN, Hilden, Germany), following previously described procedures [67]. Negative controls from all experimental steps were included to monitor external contamination. Library preparation and sequencing were performed at the Institute of Clinical Molecular Biology, Kiel University, Germany. Libraries for the sequencing runs were generated with a modified protocol for Illumina multiplex sequencing [68, 69]. The sequencing was carried out on the Illumina HiSeq 2000 and 2500 platform by 2 × 101 cycles using the HiSeq v3 chemistry and the manufacturer's protocol for multiplex sequencing. For details to the metagenomic shotgun datasets please refer to [Data S1_Illumina shotgun datasets](#).

Bioinformatic analysis of the Illumina datasets

The metagenomic shotgun datasets were subjected to a bioinformatic pipeline to identify the major animal and plant dietary composition of the Iceman's last meals. The paired-end Illumina reads (101 bp length) were first quality-checked using FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) and then subjected to adaptor removal and read merging using SeqPrep (<https://github.com/jstjohn/SeqPrep>) ("SeqPrep -f forward-read-fastqfile -r reverse-read-fastqfile -1 forward-read-output-fastqfile -2 reverse-read-output-fastqfile -L 15 -A10 "AGATCGGAAGAGCACACGTCTGAA" -B "AGATCGGAAGAGCGTCGTGTAGGG" -s merged-fastq-file). Approximately 60%–90% of reads were merged (see [Data S1_Illumina shotgun datasets](#)). To obtain a first overview of the taxonomic composition of selected samples from the different gastrointestinal tract contents and of the muscle tissue control sample, we performed a sequence similarity search of the metagenomic reads using RAPSearch2 [38] and the complete NCBI-nr database [70]. A stringent minimum bitscore filter (min bitscore 70) was applied to reduce random hits and the results were taxonomically assigned using MEGAN6 [40] with the lowest common ancestor (LCA) of hits within 10% of the bitscore of the best hits and a minimum number of 100 reads that a taxon must obtain. Finally, the data was visualized using the Krona metagenomic visualization tool [39] providing a first taxonomic overview on the bacterial and eukaryotic reads. In the subsequent step, we focused on the eukaryotic reads aiming at identifying highly abundant animal and plant species possibly resembling the Iceman's diet. We first aligned all metagenomic reads against all currently available full mitochondrial and chloroplast genomes of the NCBI database [70] using BWA [41] with default parameters. Subsequently, to obtain a taxonomic overview of the mapped reads and to identify and filter for the most prevalent taxonomic groups in the samples, we performed a sequence similarity search using

blastn [42] with default parameters and the complete NCBI-nt database as reference database [70]. Blast results were taxonomically assigned using MEGAN6 [40] with a Min percent value of 0.01 for the reads mapped against the mitochondrial genomes and with a Min percent value of 0.5 for the reads mapped against the chloroplast genomes (see Figure S2).

To further confirm our previous taxonomic assignments and to understand if the extracted reads are of ancient origin or if they display modern contamination, we reconstructed the plastid genomes of selected animal and plant species and performed phylogenetic assignments and DNA damage pattern analysis. Initially, for the plastid genome reconstruction, we extracted all reads of all intestinal content shotgun datasets (See Data S1_ Illumina shotgun datasets, both UDG-treated and non-UDG treated) that were previously taxonomically assigned to the four dominant animal and plant (sub)families *Caprinae Cervinae*, *Dennstaedtiaceae* and *Triticeae* (see Data S1_ Mapping metagenomic reads). Next, we selected for the alignment of each (sub)family reads the modern reference sequence based on the dominant species detected in the previous taxonomic assignment: the mitochondrial genome of *Capra ibex* (NC_020623) for the *Caprinae* reads, the mitochondrial genome of *Cervus elaphus* (AB245427) for the *Cervinae* reads, and the chloroplast genome of *Pteridium aquilinum* subsp. *aquilinum* (HM535629) for the *Dennstaedtiaceae* reads. However, the *Triticeae* reads have been only taxonomically assigned down to the genus *Triticum*. Thus, to obtain an appropriate modern reference for the alignment, we mapped the *Triticeae* reads against selected chloroplast genomes of domesticated and wild species in the genus *Triticum* using BWA [41] (with default parameters) implemented in the program FastQ Screen [71]. Thereby obtained FastQ Screen results indicate that the *Triticeae* reads preferentially map to both *Triticum monococcum* (KC912690) and *Triticum urartu* (KC912693) having the most specific reads to these two *Triticum* species (see Data S1_ FastQScreen Triticeae reads).

The previously extracted reads (both UDG-treated and non-UDG treated) of the four dominant animal and plant (sub)families *Caprinae Cervinae*, *Dennstaedtiaceae*, and *Triticeae* were then re-mapped against the respective reference plastid genomes using BWA [41] and increased the mapping sensitivity parameters (-l 1000, -o 1, -n 0.03). The obtained mapping statistics are summarized in the (see Data S1_ Mapping metagenomic reads). Resulting SAM files were filtered for a minimum mapping quality of 30, converted to BAM files, sorted, and indexed with SAMtools [43]. Duplicates were removed using MarkDuplicates from Picard tools (version 115, <http://broadinstitute.github.io/picard/>). A consensus FASTA sequence was generated for each sample using the ANGSD tool [44] and making a majority consensus call from overlapping reads (with the arguments -doFasta2 -doCounts1).

The reconstructed plastid genome sequences were subjected to comparative sequence and phylogenetic assignment. At first, a comparative plastid genome dataset was created. Nucleic acid sequences of modern plastid genomes that are closely related to the consensus sequences were retrieved from the public database using the BLAST search tool NCBI tblastn [42]. A multiple alignment was obtained by using the MAFFT multiple sequence alignment program [45]. The automatically inferred alignment was manually refined by using the sequence editor implemented in the ARB software package [46]. Phylogenetic analyses were performed by applying the maximum-likelihood method [PhyML [72] with the JTT substitution model]. For all datasets a stringent filter approach was applied to include into the phylogenetic assignment only position present in all sequences (please refer to Figure S3 for details to the number of informative position used).

To assess the nucleotide misincorporation patterns along the DNA fragments, we performed a mapDamage analysis [22] using all non-UDG treated reads of the Iceman intestine contents mapped against the above mentioned reference genomes. Mapping and filtering has been done as described for the preparation of the consensus sequences for the phylogenetic analysis (see Data S1). Thereby generated SAM files and the corresponding reads mapped against the reference genomes were used for the mapDamage analysis (see Figure S3D).

One limitation of the previous applied taxonomic assignment approach is the currently still limited availability of full plant and animal plastid genomes. Especially the chloroplast genomes deposited in the NCBI database represent only a small percentage of the possible plant diets present in the European Alps during the Iceman's time. Therefore, we decided to perform a comparative sequence analysis of selected shotgun read datasets against the DNA barcodes deposited in the BOLD system database (<http://www.boldsystems.org/>) [51]. The DNA barcodes deposited at the BOLD database comprise fragments of the cytochrome c oxidase I (*COI*) gene, the ribulose biphosphate carboxylase (*rbcL*) gene and the maturase K (*matK*) gene and the number of deposited species by far exceeds the number of plastid genomes in the NCBI database. The comparative analyses were performed against the *COI* fragments deposited in the groups *Mammalia*, *Actinopterygii*, *Astacidae*, *Nematoda*, and *Platyhelminthes* and against the *rbcL* and *matK* fragments in the groups *Magnoliophyta*, *Pinophyta*, *Bryophyta*, and *Pteridophyta*. First, we performed a sequence similarity search with the reads of selected shotgun datasets using blastn [42] with default parameters against the DNA barcodes extracted from the BOLD database. In the analysis four Iceman intestinal content datasets (B0626, B0621, C1824, B0625) were included, together with the Iceman muscle tissue (C0004) and Extraction Blank (B0629) as control datasets. All hits from the previous step with minimum bitscore 70, min length 15, 80% alignment length, 95% identity were further subjected to a second sequence similarity search using blastn [42] with default parameters and the complete NCBI-nt database as reference database. Blast results were taxonomically assigned using MEGAN6 [40] with a Min percent value of 0.01 (see Figure S4A).

Animal mitochondrial genome enrichment in the Iceman's stomach content

Ancient DNA extraction and library preparation were conducted in the dedicated ancient DNA laboratory at EURAC research, Institute for Mummy Studies, Bolzano. The DNA was extracted from stomach contents using in-solution silica based protocol for ancient DNA [73]. The extracted DNA was prepared for Illumina sequencing by blunt-end repair and ligation of universal Illumina adapters (31). The library was shipped at -20°C to Trinity College Dublin and Illumina barcoded at the 5' end there [74]. A custom RNA capture produced from reference domesticated mitochondrial genomes by MYcroarray (Mycroarray, 5692 Plymouth Road, Ann Arbor, MI

48105) was used to enrich the indexed library for mitochondrial genomes (following the manufacturer's protocol). The captured library was sequenced on the Illumina MiSeq platform at TrinSeq St James's Hospital Dublin (65 bp single-end sequencing kit). Data were quality assessed with fastqc, sequencing produced 4,905,473 raw reads, of which 95.5% passed adaptor trimming by AdapterRemoval [47]. To provide an overview of which mammalian DNA sequences were captured, the recovered reads were mapped to an array of candidate mammalian genomes using FastqScreen (https://www.bioinformatics.babraham.ac.uk/projects/fastq_screen/) (see Data S1_FastQScreen enriched reads) and bwa aligner (arguments: `aln -l 1000 -n 0.01`). Mapping sequence data showed that the captured library contained reads specific for red deer (*Cervus elaphus*, reference accession code: AB245427), ibex (*Capra ibex*, reference accession code: NC_020623) and human (reference accession code: NC_012920.1). Mapped reads were further analyzed for these three species, SAMtools [45] filtered out reads below mapping quality 30. DeDup (<https://github.com/apeltzer/DeDup>) removed duplicate reads (see Data S1). mapDamage displayed the presence of lesions associated with ancient DNA [22] (see Figure S3D), showing that on 10% of fragments had deamination lesions at the 5' end of fragments. To investigate variants in the mapped mitochondrial data, GATK UnifiedCaller created *vcf* files from the bam files and variants were quantified with MultiVcfAnalyser, genotypes with a quality below 30 were filtered [48]. To further reduce the potentially confounding data from cross mapping or misincorporation of deaminated positions, only SNPs with a consensus above 80% were considered with respect to the overlapping reads (see Data S1_Variants enriched reads). This filtering strategy effectively removed confounding SNPs as demonstrated by the exact match to the Iceman's mitochondrial haplotype defining SNPs [75].

Polymerase chain reaction (PCR)-based analyses

To further confirm our metagenomic findings in the Iceman intestinal content shotgun datasets we additionally performed a PCR-based analysis with selected DNA extracts targeting small DNA fragments of animal and plant barcodes (see Key Resources Table). To assess the animal diet of the Iceman, we amplified the fragments of two different mitochondrial loci, the 12S rRNA gene (12S rRNA) and the cytochrome B gene (*cytB*). For the *cytB* gene we applied a previously published PCR assay specifically designed to target different animal species [35]. In parallel, this assay excludes the amplification of human background DNA. Furthermore, we designed for this study a second PCR assay targeting a smaller fragment of the 12S rRNA gene of different animal species, including the human 12S rRNA gene variant. To further investigate via PCR the plant diet of the Iceman we amplified in the intestinal content DNA extracts the P6 loop of the chloroplast *trnL* (UAA) intron [37] and a 153 bp fragment of the chloroplast gene coding for the large subunit of the ribulose biphosphate carboxylase (*rbcL*) [36]. To test for possible external contaminations, we included in all PCR assays the extraction blanks and two negative PCR controls. In addition, we included in the plant PCR assays an Iceman lung tissue sample to test for possibly inhaled pollen background. All PCR assays have been performed in the ancient DNA laboratory of the Eurac research Institute for Mummy Studies conducted in 50 µl volume using the Amplitaq Gold mastermix 360, 1 mM primers and 5 µl DNA template. Template DNA was initially denatured at 95°C for 5 min followed by 40 cycles of denaturation (95°C, 45 s), primer annealing (55°C, 45 s) and elongation (72°C, 45 s), with 4 min of final extension at 72°C. Only for the 12S rRNA gene PCR assay 30 cycles instead of 40 cycles were applied. The first PCR was followed by a second PCR using 1:50 dilution of the first PCR product and 10 to 15 additional cycles of amplification with barcoded PCR primers [76]. The barcoded Primers consisted of the gene-specific PCR primer sequences tagged with the sequencing adapters for GS FLX Titanium chemistry, including (5'-3'): Titanium adaptor (454 Life Sciences), a sample specific ten-base multiplex identifiers (MIDs on the reverse primer), a linker sequence ("TC" for forward primer, "CA" for reverse primer), and the gene-specific PCR primer. Amplicons were sequenced from the reverse side on a GS FLX instrument using Titanium chemistry (454 Life Sciences). For details to the amplicon datasets please refer to Data S1_454 Amplicon datasets. Sequencing reads were de-multiplexed and filtered using the mothur software package [49]. To obtain a taxonomic overview of the reads, we performed a sequence similarity search using blastn (40) with default parameters and the complete NCBI-nt database as reference database. The results were taxonomically assigned using MEGAN6 [40] with the lowest common ancestor (LCA) of hits within 10% of the bitscore of the best hits and a minimum number of 2 reads that a taxon must obtain (see Figure S4).

Proteomic analysis of the Iceman stomach content

We studied the proteome of three distinct stomach content samples (1051L, 1051LP & 1054LP) by liquid chromatography-mass spectrometry (LC-MS) proteomics using Orbitrap and QExactive technologies. For methodological details to the sample preparation, the LC-MS approach and the mass spectrometry data analysis please refer to the publication of Maixner et al. [11]. Briefly, each stomach content sample was solubilized, tryptic digested and fractionated using 1D SDS-PAGE or OffGEL isoelectric focusing (OGE) and then analyzed by high-mass accuracy Orbitrap or QExactive mass spectrometry instruments interfaced with nanospray liquid chromatography. Peptide identities were determined using the Trans-Proteomic Pipeline (TPP) software tool suite to define proteins contained in the stomach content samples [50].

The protein database used in the comparative analysis contained amino acid sequences constructed from multiple species. 75,649 protein sequences were to *Homo sapiens* (UniProt reference proteome, release 2012_10 ftp.uniprot.org/pub/databases/uniprot/previous_releases/release-2012_10/relnotes.txt) supplemented with sequences translated directly from the Iceman's genome [34, 58]. Additionally, 936 and 1,278 *Cervus elaphus* protein sequences were obtained from NCBI and UniProt, respectively (obtained November 17, 2015). 30,155 *Capra hircus* protein sequences were obtained from NCBI (October 19, 2015) as the closest surrogate for the ibex proteome. The 1,011 available *Triticum monococcum* protein sequences from UniProt were supplemented with 95,051 *Triticum aestivum* sequences from UniProt (release 2015_11) to provide a complete proteome representing *Triticinae*. 28,356 open reading frames of at least 30 amino acids were obtained from a previously published *Pteridium aquilinum* transcriptome

[77]. An additional 67 protein sequences to common laboratory contaminants were also added. An equal number of shuffled decoy sequences were generated to perform database search validation, as described previously [11].

The identified proteins (167), evaluated by statistical analysis using the TPP to 1% false discovery rate, were further taxonomically screened for homologous peptides and subsequently filtered for peptide sequences that can be unambiguously assigned to certain plant and animal families. To taxonomically assign the retrieved peptides based on the lowest common ancestor we performed a BLAST search [42] against the NCBI nr database and further analyzed the BLAST hits with the MEGAN6 software package [40]. In total, 20 of the 167 proteins were unambiguously identified with peptides assigned to the families *Triticeae*, *Caprinae*, and *Cervinae* (see [Data S2](#)_ Proteomics).

Lipid analysis of the Iceman stomach content

During the preliminary inspection of the Iceman's stomach content, two samples were isolated, described as "lipid accumulation" and "lipid/protein mix." Both samples were inhomogeneous, and multiple aliquots (a few milligrams each) were taken for lipid analysis. Aliquots were extracted using chloroform-methanol/based liquid-liquid extraction and analysis using an unbiased approach with reverse phase liquid chromatography separation and quadrupole time-of-flight (QToF) tandem mass spectrometry analysis (RP-LC/MSMS). Lipid analyses of aliquots from the Iceman stomach content were undertaken independently at two laboratories in Singapore and Seattle, USA. The analytical methods were as follows:

Singapore analytical method

Sample Preparation: Portions of two Iceman stomach samples and fresh animal tissue samples were dissected with a scalpel on an aluminum foil-covered ice pack, and the weights were noted: 1) Iceman Lipid accumulation 1: 6.9 mg, 2) Iceman Lipid/protein mix 1: 4.5 mg, 3) Capra ibex muscle tissue: 17.9 mg, 4) Capra ibex adipose tissue: 8.2mg, 5) Capra ibex skin: 8.6mg, 6) Cervus elaphus muscle tissue: 18.3 mg, 7) Cervus elaphus fat tissue: 15.0 mg, 8) Capra hircus cheese: 17.8mg, 9) Capra hircus milk: 27.5mg. For details to the modern reference samples used in this study please refer to Table S12. Lipids were extracted by a modified Bligh and Dyer extraction method [78]. The tissue pieces were transferred to glass homogenization tubes, and 300 μ L of methanol were added. Samples were homogenized using a benchtop homogenizer, and the homogenizer probe was rinsed with an extra 300 μ L methanol. 300 μ L of chloroform were added to the homogenates (final volume 900 μ L of chloroform/methanol 1/2 v/v). The homogenizing rotor was carefully rinsed between each sample. A tube containing methanol only was used as negative control. The bottom organic phases were recovered and transfer to clean tubes and the solvent was dried under nitrogen gas. Dried lipid extracts were kept at -80°C , until LC/MS analysis for which they were re-suspended in 200 μ L chloroform/methanol (1/1 v/v). To determine the approx. lipid amount (w/w) three stomach biopsies were first dried at 105°C for 24h and weighed. Then, the lipids were extracted by the modified Bligh and Dyer extraction method (described above) and the extracts were dried at 105°C for 24h and weighed.

LC/MS: Chromatography conditions were optimized to enable efficient separation of triacylglycerols (TAGs) by reverse phase separation. LC/MS experiments were undertaken using an Agilent 1200 series HPLC-Chip system connected to Agilent 6540 Q-ToF. An Agilent C18 chip was used (75 μm (i.d.) x 43 mm length, 5 μm particle size, 300 \AA pore size) with a binary solvent system at a flow rate of 0.3 $\mu\text{L}/\text{min}$. Solvent A was acetonitrile/water 40/60 v/v containing 5mM ammonium acetate and 0.1% acetic acid, and solvent B was isopropanol/acetonitrile 90/10 v/v containing 5mM ammonium acetate and 0.1% acetic acid. The gradient was as follows: 0%B for 1.5min, increase to 50%B in 2.0min, increase to 100% B in 18min, stay at 100% B for 10min, decrease to 0%B in 0.1min, stay at 0%B for 3.4min, total runtime: 35min. The Agilent 6540 Q-ToF spectrometer was operated in positive ion mode; electrospray voltage was set to 1450 V (Vcap), temperature 300°C , drying gas 4L/min, fragmentor voltage 150 V. The instrument was operated with MS acquisition rate of 1 spectra/sec. Injection volume was 1 to 2 μL . To monitor sample LC carryover and cross-contamination, solvent blanks were run between samples.

Data Analyses: LC/MS data files were processed with the MassHunter Qualitative Analysis Software Version B.05.01. The data were manually curated to identify triglyceride ions ($[\text{M}+\text{NH}_4]^+$ adducts) based on retention time and accurate mass measurement with mass accuracy $< 5\text{ppm}$. Intensities of the $[\text{M}+\text{NH}_4]^+$ adduct mono-isotopic peaks were used for calculating the relative abundance of triglyceride molecular species. TAG have been successfully used in the identification of food remains in an archaeological context [26, 79]. Therefore, we decided to compare the relative abundance of TAG between the Iceman's stomach samples and with contemporary animal samples (See [Data S2](#)_Modern comparative samples). Following the genomics results, *C. ibex* and *C. elaphus* meat samples were obtained from local hunters. In addition, and in view of the interest regarding the question of dairy consumption in early humans, samples of *C. hircus* milk and cheese were also sourced. To better compare TAG distribution between samples, Mirabaud et al. [26] made use of two parameters: the average carbon number (M) and the distribution factor (DF), which can be calculated by the following formulae:

$$M = \frac{\sum(P_i C_i)}{\sum P_i}$$

$$DF = \frac{\sqrt{[(C_i - M)^2 C_i P_i]}}{\sum P_i}$$

where P_i is the relative abundance of a given TAG (in percentage of total TAG) and C_i is the total number of carbon atoms in the fatty acyl chains.

Seattle analytical method

Sample Preparation: Portions of two Iceman samples and four fresh male tissue samples were dissected with a razor on an aluminum-foil covered block on top of dry ice, and the weights were noted: 1) Iceman Lipid accumulation 5: 73 mg, 2) Iceman Lipid-protein mix 5: 70 mg, 3) 0065206-19 adipose: 182 mg, 4) 0065251-10 adipose: 135 mg, 5) 0065251-29 skin (full thickness): 209 mg, 6) 0065206-09 skin (full thickness): 184 mg. The tissue pieces were transferred to homogenization tubes containing 2.38 mm metal beads, and 1 mL methanol. To serve as negative controls, two tubes containing methanol only were carried through each step of sample preparation. Following bead beating, BCA protein estimation was performed on the soluble portion of each lysate. The fresh tissue samples each contained between 500–2000 μg of soluble protein, and, surprisingly, each of the two Iceman samples contained roughly 500 μg soluble protein. The lysates were transferred to silanized 13x100 glass vials, and total lipids were extracted with chloroform/methanol using a modified “Folch” extraction protocol [80]. The combined aqueous upper phase and interphase (containing protein precipitates) from each sample were dried and processed by multi-dimensional fractionation for proteomics analyses in Seattle (see above). The lower organic phases were dried under nitrogen and stored at -30°C . The visual appearance of the dried lipid extracts differed remarkably: Iceman extracts appeared as a dry brown powder, and the fresh tissue extracts appeared as a viscous yellow liquid. For lipid LC/MS, 8 mL chloroform/methanol (2:1 v/v) was added to each extract, and 5 μL was used for injection. The extract was also diluted 100-fold and 5 μL was again used for injection. Additionally, neutral lipids were obtained from the total lipid extract by solid-phase extraction (SPE) following a previously published method [81]. Briefly, 100 μL of the total lipid extract was evaporated under N_2 gas, re-dissolved in 1 mL isooctane/ethyl acetate (75/25 v/v), sonicated, and then loaded onto 100 mg Biotage Isoolute XL silica cartridges. Neutral lipids were collected from the flow-through, and further washed and collected from the column with isooctane/ethyl acetate (75/25 v/v). Eluants were evaporated under nitrogen gas, dissolved in 1 mL chloroform/methanol (2:1 v/v), and 5 μL was used for injection. Additionally, a 10-fold dilution was also used for injection.

LC/MS: Chromatography conditions were optimized to enable efficient separation of triacylglycerols (TAGs). An Agilent 1290 Infinity LC with a 2.1 mm (i.d.) x 100 mm, 3.5 μm Agilent ZORBAX Eclipse XDB-C18 Narrow Bore Rapid Resolution column heated to 30°C was used with a binary solvent system and a flow rate of 500 $\mu\text{L}/\text{min}$. The system was equilibrated with 80% of solvent A (5 mM ammonium formate in methanol/water (99:1 v/v)) and 20% solvent B (5 mM ammonium formate in isopropanol/water (99:1 v/v)). A 5 μL injection volume of the lipid extract (in 2:1 chloroform/methanol) was applied to the column, followed by a 30.0-min linear gradient to 20% solvent A/80% solvent B, and held for 2.5 min. An Agilent 6530 Q-ToF equipped with an Agilent JetStream ESI source was used for accurate mass analysis of the LC eluent. Positive ion mass spectra data were acquired in 2 GHz Extended Dynamic Range mode at a rate of 1.02 spectra/s, and data was collected as profiled spectra over a mass range of 150 to 1500 m/z . For MS/MS analyses, a fixed collision energy of 40 V was used for fragmentation. To monitor sample LC carryover and cross-contamination, several solvent blanks were run between samples, and sample order was considered carefully.

Data Analyses: LC/MS data files were processed with the MassHunter Qualitative Analysis Software version B.03.01. Molecular features (MFs) were extracted from the raw MS1 data using the Molecular Feature Extraction (MFE) algorithm with a peak filter of $\geq 10,000$ counts. The resulting MFs were then identified with MassHunter by searching a custom database containing 690 TAG ions corresponding to $[\text{M}+\text{Na}]^+$ and $[\text{M}+\text{NH}_4]^+$ adducts, and 99 $[\text{M}+\text{NH}_4]^+$ cholesterol ester ions, using a ± 5 ppm search window. For quantitative purposes, lipid groups were compared using the $[\text{M}+\text{NH}_4]^+$ molecular ion height. See [Data S2](#) for a summary of the lipid analysis results.

Atomic force microscopy (AFM) and Raman spectroscopy

Meat fiber assemblies within the Iceman food bolus samples were identified with an inverted optical microscope (Axiovert 135, Zeiss, Oberkochen, Germany). Subsequently, the coarse topography of the putative meat fibers was analyzed by laser scanning microscope (LSM) images (VK 8710, Keyence, Osaka, Japan) while high-resolution images of defined sample areas were taken with a NanoWizard-II AFM (JPK Instruments, Berlin, Germany). The AFM was operated in the intermittent contact mode and silicon cantilevers (BS Tap 300, Budget Sensors, Redding, USA) with nominal spring constants of 40 N/m, resonance frequencies of 300 kHz, and tip radii of 10 nm were used. All images were taken at lab conditions (22°C and about 45% relative humidity).

To assess the molecular composition of the samples, Raman spectra were taken with a confocal Raman microscope (WITec alpha 300 R, WITec GmbH, Ulm, Germany). The Raman system was operated with a laser (frequency doubled Nd:YAG-laser with an excitation wavelength of 532 nm) at 1.0 mW laser power, to avoid sample damage (photodegradation). The backscattered light from the sample was detected with a vacuum sealed, high-sensitivity, back-illuminated CCD camera cooled to -56°C . The spectrometer was equipped with a 600 g/mm grating enabling a spectral resolution of approximately 3 cm^{-1} per CCD-pixel. Single spectra of each sample are average spectra of 10 accumulations with up to 180 s of integration time. Due to the confocal setup of the microscope and the objectives used, Raman spectra were collected from a sample area with approximately 400 nm diameter and a focal depth of about 1 μm .

QUANTIFICATION AND STATISTICAL ANALYSIS

Phylogenetic analyses were performed by applying the maximum-likelihood method [PhyML [58] with the JTT substitution model] implemented in the ARB software package [46]. For all datasets a stringent filter approach was applied to include into the phylogenetic assignment only position present in all sequences.

DATA AND SOFTWARE AVAILABILITY

The Illumina datasets are available from the European Nucleotide Archive (ENA) under accession number ENA: ERP012908. The enriched Illumina dataset has been deposited in ENA under the accession number ENA: PRJEB26465. The 454 amplicon datasets has been deposited in ENA under the accession number ENA: PRJEB26365. The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE partner repository [82] with the dataset identifier PXD009565.

ADDITIONAL RESOURCES

Additional metadata for the Illumina shotgun datasets are provided under the following link: <http://cube.univie.ac.at/supplements/iceman>.